Research Article

Primary cutaneous B-cell lymphomas: our experience with 22 cases

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Abstract

Background. Primary cutaneous B cell-lymphomas (PCBCL) are a group of lymphomas with clinical, prognostic, and therapeutic characteristics differing from nodal lymphomas. These facts determined that, in the last five years, they were grouped as an independent entity.
Objective. To present the experience of our Dermatology Department as far as diagnosis, management and follow-up of primary cutaneous B-cell lymphomas are concerned, during the last eleven years
Design. Descriptive and retrospective study.
Materials and methods. 22 patients with diagnosis of primary cutaneous B-cell lymphomas were studied between 1995 and 2006, at the Dermatology Department, Policlínico Bancario.
Results. Our experience comprised 22 patients, of which 13 were follicle center B-cell lymphomas (59 percent), 4 were marginal zone B-cell lymphomas (18.18 percent), 2 were diffuse large B-cell lymphomas (leg-type), 2 were diffuse large B-cell lymphomas (other), and only 1 case was a mantle cell lymphoma.
Conclusion. The last PCBCL classification takes into account the clinical, therapeutic, histopathologic, and genetic aspects. This classification optimizes dermatologist management of this pathology and enhances their relevance within the interdisciplinary group in charge of the follow-up. However, a permanent updating must be considered that might include new changes in a not distant future (Dermatol Argent 2008;14(1):35-45).

Key words: *primary cutaneous B-cell lymphoma, PCBCL.*

Introduction

Primary cutaneous B-cell lymphomas (PCBCL) are clonal B-cell proliferations perfectly separated from nodal lymphomas and their secondary cutaneous involvement. They comprise a group with clear clinical, therapeutic, and prognosis differences. In addition, because they show different chromosome translocations, variable oncogenes expression, and specific viral sequences, primary cutaneous lymphomas comprise a clinically and biologically independent entity.
These concepts were clarified only recently, since in the last 50 years multiple classifications were proposed, not distinguishing them from nodal lymphomas.
In order to speak about a true primary cutaneous lymphoma, we must ascertain the absence of extracutaneous involvement at the time of diagnosis.
There are clinical, histopathological, and immunohistochemical guidelines and supplementary studies to establish diagnosis and staging of B-cell lymphoma.[1-5]

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The EORTC (European Organization for Research and Treatment of Cancer) classification promotes an organ-specific classification, *i.e.* specific for cutaneous lymphomas, and incorporates the terms indolent and aggressive in its lymphoma division, thus including a prognostic feature.[6] In spite of having been diversely criticized, this classification was adopted by dermatologists worldwide and was the foundation for the unified classification known as WHO-EORTC (**Table 1**).[7-11]

Materials and method

An observational, descriptive, and retrospective study was conducted, including 22 patients with diagnosis of primary cutaneous B-cell lymphoma, during the 1995-2006 period, at the Dermatology Department of Policlínico Bancario.
Frequency was assessed according to patient age and gender, types of lymphoma, presentation, location, and treatment, as well as 5-year survival.
Diagnosis was reached according to clinical examination, histopathology, immunohistochemistry, and the study of light chains, in all cases.[12,13]
Supplementary studies were performed every 6 months, to rule out systemic involvement, with 5-year follow-up of patients.

Results

The 22 cases of primary cutaneous B-cell lymphomas compared to 38 cases of primary T-cell lymphomas seen during the same period of time, amount to 36 percent of the total of observed lymphomas, a percentage somewhat higher than the internationally stated (30 percent), possibly due to the elderly population attended at our institution.
From the total 22 cases of our experience, 13 referred to follicle center lymphomas (59 percent), 11 in male and 2 in female patients; 4 were marginal zone lymphomas (18.18 percent), with 3:1 females dominance; 2 cases were large cell lymphoma of the leg (9.09 percent), 1 male and 1 female; 2 male cases were other large cell lymphoma (9.09 percent); and only 1 male had primary cutaneous mantle cell lymphoma (4.54 p) (**Table 2** and **Graphic 1**).
Age distribution for the total sample and for each variant was as follows (in years):

- Total patients: mean 59.50 (range: 39-91).
- Follicle center lymphomas: 61.6 (48-91).
- Marginal lymphomas: 52 (39-62).
- Large leg-type lymphomas: 73.5 (71-76).
- Other large cell lymphomas: 50 (41-59).
- The mantle lymphoma patient was 55 years old.

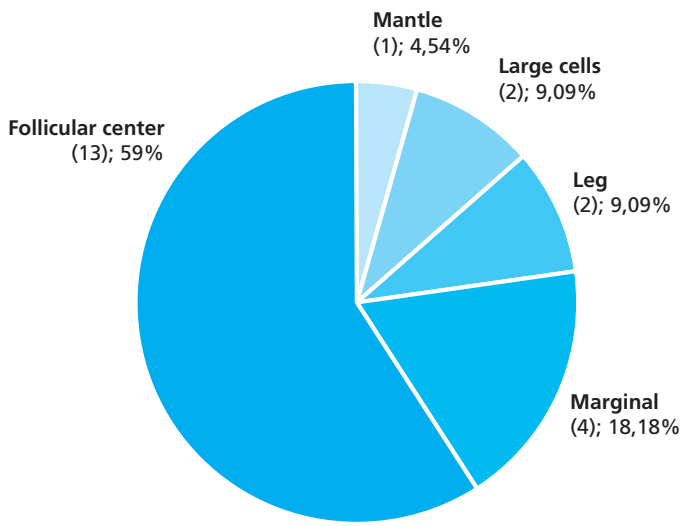
TABLE 1. CLASSIFICATION OF CUTANEOUS B-CELL LYMPHOMAS (WHO-EORTC).

• Primary cutaneous marginal zone B-cell lymphoma.
• Primary cutaneous follicular center B-cell lymphoma.
• Primary cutaneous diffuse large B-cell lymphoma, leg-type.
• Other primary cutaneous diffuse large B-cell lymphomas (includes intravascular large B-cell lymphoma).

TABLE 2. CASE HISTORY.

Case	Type	Gender	Age	Clinic	Location	Treat.	SV
1	FC	M	48	T	Scalp	RT	Yes
2	FC	M	60	T	Right temporal	RT	Yes
3	FC	M	66	T	Left temporal	RT	Yes
4	FC	M	48	T	Scalp	RT	Yes
5	FC	M	63	T	Face	RT	Yes
6	FC	F	69	T	Face	RT+CT	No
7	FC	M	65	T(4)	Face, neck, back, breast	RT	Yes
8	FC	M	61	T	Back	RT	Yes
9	FC	M	71	T(2)	Back (recurrent)	RT	Yes
10	FC	M	55	Multiple	Back (Crosti-type)	RT+CT	Yes
11	FC	M	52	Multiple	Flank (Crosti-type)	RT	Yes
12	FC	M	53	Multiple	Abdomen	RT	Yes
13	FC	F	91	T(2)	Back	S	No
14	Marg.	M	48	T, nodule	Thighs	S	Yes
15	Marg.	F	39	Plaque	Forearm	RT	Yes
16	Marg.	F	62	T	Face	S	Yes
17	Marg.	F	59	T	Scalp	RT	Yes
18	LCL	F	76	Plaque	Leg	RT+CT	No
19	LCL	M	71	T	Foot	RT	Yes
20	IVLC	M	59	Plaque	Pectoral	RT+CT	No
21	PlasLC	M	41	T(3)	Arm, thorax, neck	CT	No
22	Mantle	M	55	T(3), panniculitis	Thigh	RT	Yes

FC: follicular center. Marg: marginal. P: plasmocytoma. LCL: large cells leg type. IVLC: intravascular large cells. PlasLC: plasmoblastic large cells. T: tumor. SV: 5-year survival. RT: radiotherapy. CT: chemotherapy. S: surgery.



Graphic 1. *Lymphoma distribution [(amount of patients); percentage].*

Five-year survival percentage was assessed throughout the sample, because that follow-up period was left to include the experience cases:

- For the total sample of 22 patients, it amounted to 77 percent (5 died, but the 91-year-old patient died for causes not related to the disease).
- For each lymphoma group: follicle center, 85 percent; marginal, 100 percent; leg type, 50 percent; other large cell, 0 percent; mantle, 100 percent.

Discussion

The results were analyzed according to the WHO-EORTC classification, and our clinical, histopathological and immunohistochemical findings were compared to the reviewed literature on the subject matter. A notable coincidence was found, thus we unified concepts in the development of each variant.

Primary cutaneous follicular center B-cell lymphoma

These are follicular center cell neoplasms with variable amount of centrocytes (small and large cleaved follicular center cells) and centroblasts (non-cleaved large follicular center cells with prominent nucleus) of follicular, follicular and diffuse, or diffuse growth.[14]

Clinically, they appear as red papules, plaques, or nodules, isolated or aggregated, slow growing, usually in the head, neck, and trunk[15] (**Figure 1**). Of all patients with follicular center lymphomas (13), the most frequent clinical presentation was a tumor, with the following distribution: six in the head, six in the trunk, and one with multiple locations in face, neck, back, and breast. Two patients (cases 10 and 11) had the "Crosti's reticulo-histiocytoma of the back," with papule lesions and plaques distributed over an erythematous base, preceding the tumor in month or years[16-19] (**Figure 2**). Histologically, these lymphomas show a nodular or diffuse pattern preserving epidermis. In the early stage centrocytes, centroblasts, reactive cells and remaining reactive follicular centers are identified, without mantle cells;[20,21] and in the tumor stage, there is an increase in the number and size of neoplastic cells.[15,16,22-24] Fast growing lymphomas contain large follicular center cells, centroblasts, large centrocytes, multilobular cells, and immunoblasts. In all cases, there was marked fibroblast reaction.

Immunohistochemistry shows positive results for CD19, CD20, CD22, CD79a, CD10, Bcl-6, and negative for CD5.[18,19,25-29] Surface immunoglobulin (sIg) monoclonality was confirmed.[30]

Figure 1. *Follicular center B-cell lymphoma. Classic clinic aspect (Case 2).*

Figure 2. *Crosti's reticulo-histiocytoma of the back (Case 10).*

These lymphomas show Ig clone rearrangement, but no t(14;18) translocation. No expression of Bcl-2 protein is identified.

Additionally, somatic hypermutation of the heavy and light chain variable gene is described, thus indicating their follicular center origin.[31,32]

Independently on the nodular or diffuse growth pattern, and isolated or multiple appearance, 5-year survival is 95 percent.[21,22,33]

Our patients showed similar survival to the mentioned statistical data: one of the two deceased patients died because of the lymphoma (case 6) and the other (case 13) due to natural causes.

A recent study suggests that the patients with marked expression of

Bcl-2 with diffuse large cell infiltrate may have a poorer prognosis.[34]

We applied radiotherapy in the case of localized or few lesions, as suggested by the reviewed publications.[22,23,35-38] In the presence of tumor recurrence, the recommended treatment is also radiotherapy. Anthracycline is used as treatment of extended lesions or extracutaneous progression.[20,22]

We used radiotherapy in 12 out of 13 cases. In case 6, we supplemented treatment with chemotherapy. Case 10 showed post-radiotherapy tumor recurrence, and required surgical excision of the lesion. Only case 13 received surgical treatment exclusively, because the patient was incapable of being moved.

Rituximab was used systemically and intralesionally in small series of patients, with good results, but its long-term effect has not been assessed yet.[39,40]

Figure 3. *Marginal zone B-cell lymphoma B, with positive Borrelia burgdorferi serology (Case 15).*

Primary cutaneous marginal zone B-cell lymphoma

This group includes different types of lymphoma receiving diverse denominations, namely: MALT-SALT, lympho-plasmocytoid, plasmocyte, lymphoid follicular hyperplasia, immunocytoma, and plasmocytoma.

The initial immunocytoma denomination was adopted by EORTC from the KIEL classification, because of its supposed plasmocytoid differenciation.[1,41] These lymphomas actually derive from the marginal zone of the germinal center. They show wide histological variety of small B cells (centrocyte-like, lympho-plasmocytoid, or plasma cells) but have always had good prognosis.[42] Therefore, and given the similarity with mucosa associated lymphoid tissues (MALT), the SALT (skin associated lymphoid tissues) denomination was suggested.

These lymphomas clearly differentiate form pseudolymphomas, and from skin involvement secondary to nodal lymphomas.[43-45]

Tick-bite transmitted *Borrelia burgdorferi* is mentioned as one etiologic factor; therefore its usual localization in exposed areas.[46,47]

These lymphomas appear as red, violaceous, solitary, or multiple, slow growing tumors located in the proximal area of limbs, gluteus, and trunk. Our experience joined four cases, whereof three had tumors, and one had plaques. As regards location, two appeared on the limbs (one on the thigh, and one on the forearm), and two on the head (one on the face [**Figure 3**] and one on the scalp). Histologically, they occur in a nodular or diffuse pattern, preserving epidermis. There was cell variability, with peripheral lympho-plasmacytoid and isolated plasma cells and central small reactive cells or reactive follicular structures, centrocytes, centroblasts, and immunoblasts. Cells with intranuclear inclusions (Dutcher bodies), characteristic of B lineage, may be found. Rarely, there is infiltration of glandular, hair, or sweat epithelia, more characteristic of node lymphomas.[41,42,46,48]

Transformation to diffuse large B-cell lymphoma is rare.

Immunophenotype of marginal zone lymphomas is: CD19, CD20, CD79a Bcl-2(+), CD5, CD10, and Bcl-6(-).[49,50] Plasma cells: CD20(-), CD138(+). Reactive germinal centers: Bcl-6, and CD10(+), Bcl-2(-).

Monoclonal cytoplasmic Ig may be found in advanced stages.

There is Ig heavy chain (IgH) clone rearrangement. Recent studies suggest the presence of t(14;18)(q32;q21) translocation involving genes codifying Ig (chromosome 14) heavy chains and MLT gene, located on chromosome 18. Also found is (3;14)(p14.1;q32) translocation involving IgH gene and FOXP1.[49,50] Gastric lymphomas have different translocations than in skin: t(11;18)(q21;q21) and t(1;14)(p22,q32).[51,52]

Cases 14 and 16 were subjected to surgical excision, and cases 15 y17, to radiotherapy; both treatments are indicated for solitary lesions.

Five-year survival is 100 percent, a similar prognosis for the 4 patients in our experience.[41,48,53]

We detected positive *B. burgdorferi* serology in case 15; thus, the patient received radiotherapy plus antibiotic treatment.[54]

In the presence of multiple lesions of primitive marginal zone B-cell lymphoma, systemic treatment with chlorambucil or interferon-alpha is suggested.[54]

Good results have been described with intralesional or systemic rituximab (anti-CD20 antibody).[55]

Case 17 is unusual in being a primary cutaneous plasmocytoma, a rare lymphoma occurring in 4 percent of extramedullary plasmocytomas.[56] This patient had a red-violet subcutaneous nodule on the scalp, of excellent prognosis, where associate multiple myeloma was ruled out[57] (**Figure 4**). Histologically noteworthy was the presence of mature plasma cells, some of them multinucleated. The immunophenotype was CD20(-), CD138(+), monoclonal for cytoplasmic Ig in plasmocytes.[58] Survival is excellent (100 percent at 5 years).The recommended treatment is radiotherapy or surgery.

Figure 4. *Primary cutaneous plasmocytoma (Case 17).*

Figure 5. *Large B-cell lymphoma of the leg (Case 18).*

Primary cutaneous diffuse large B-cell lymphoma of the leg

Dominant in patients older than 70 years, female 3-4:1, and it appears as rapidly growing red nodules or tumors in one or both lower areas of the legs; they are rare in other areas.[22,59,60] (**Figure 5**)

Case 19 showed a nodule with a particular location on a toe. These last two patients were over 70 years old. We point out that locations in head and trunk are rare.[22]

Histologically, they show diffuse infiltrates of centroblasts and immunoblasts, preserving the epidermis and extending deeply to the subcutaneous cellular tissue. There are frequent mitotic figures, and scarce stromal reaction.[22,59]

The immunophenotype was CD19, CD20, CD22, CD79a, Bcl-2(+), and CD10(-).[16,29,50,61]

There is monoclonal superficial and/or citoplasmic Ig. Protein MUM-1/IRF4 is positive with immunohistochemical techniques.[30,62]

This type of lymphoma is not associated with clone rearrangement; however, Bcl-2(+) expression is common in this group.[16,29] In some cases, this overexpression is the consequence of chromosomal amplification of Bcl-2 gene.[63]

Inactivation of tumor suppressor genes *p15* and *p16* is found in 11 and 44 percent of the cases, respectively.[64] There is chromosomal imbalance in 85 percent of tumors (18q, 7p and loss of 6q).[63,65] There may be a genetic expression profile of activated B-cells,[30] and translocation of genes *myc*, *Bcl-6* and of IgH.[66]

Prognosis of these lymphomas is unfavorable, compared to follicular center lymphomas, with greater tendency to extracutaneous dissemination.[22,59] Five-year survival is 55 percent and the presence of multiple lesions is a factor of bad prognosis.[22] In case 18, the patient died 3 years after diagnosis, in spite of receiving combined treatment (radiotherapy-chemotherapy). For patients with solitary or localized tumor, radiotherapy is indicated.[22] Said treatment was administered to case 19.

Polychemotherapy is indicated for multiple lesions, with systemic anthracycline or rituximab.[22,39,59,67]

Other primary cutaneous diffuse large B-cell lymphoma

It comprises those cases of large B-cell not included in the follicular center or leg lymphoma groups.

It includes morphologic variants such as anaplastic, plasmoblastic, or large B-cell lymphoma rich in histiocytic T cells. Some cases constitute a cutaneous manifestation of a systemic lymphoma.

Plasmoblastic lymphoma appears exclusively in HIV patients or in other immunodefficiencies.[68] Case 21 is a 41-year old HIV patient with multiple tumors located in arm, thorax, and neck, which showed torpid evolution with rapid lesion dissemination that caused the patient's death.

Large B-cell lymphomas rich in histiocytic T cells are rarely seen and are characterized by the presence of large B-cells and numerous reactive T-cells.[69,70] Those having an exaggerated reactive T-cell population have better prognosis. They are clinically similar to follicular center and marginal zone lymphomas located in the head, trunk, and limbs.

Intravascular large B-cell lymphomas may be defined as an aggregation of neoplastic large B-cells within blood vessels, and they may affect the central nervous system and the lung; the latter is associated with bad prognosis.[71] Case 20 appeared as an angiomatoid plaque that evolved to a violaceous, indurated, armor-like lesion (**Figure 6**). The lesion was reduced after irradiation; two years later, the patient developed lung involvement and died. Occasionally, lesions may acquire an early telangiectasic aspect and localize on lower area of legs and on the trunk.[71,72] Interesting cases of skin hemangioma colonization by neoplastic cells, as sole presentation symptom, are described.[73,74]

Histopatologically, there are dilated vessels in dermis and hypodermis, with proliferation of neoplastic large lymphoid cells. This cell proliferation may lead to vascular occlusion of venules, capillaries and arterioles. Extravascular aggregations of atypical cells are found in 20 percent of the cases. En case 20, these findings also appeared in the lung biopsy.

Immunophenotype was CD19, CD20, CD22, CD79a(+), monoclonal for sIg. T immunophenotype appears less frequently.

This variety of lymphomas has bad prognosis, especially if affecting central nervous system or lung. Survival at 5 years is 22 percent, but if there is only cutaneous involvement, it increases to 56 percent.[72] Chemotherapy is the treatment of choice, even in cases of exclusive cutaneous involvement.[72]

Figure 6. *Intravascular large B-cell lymphoma (Case 20).*

Figure 7. *Primary cutaneous mantle B-cell lymphoma. Panniculus clinical aspect (Case 22).*

Primary cutaneous mantle B-cell lymphoma

The current WHO-EORTC classification does not include primary cutaneous mantle B-cell lymphoma, which was stated in the EORTC classification.[7,75] Localized secondary skin lesions are not accounted for in this classification.[76]

In case 22 (**Figure 7**), the histology showed monomorphous and diffuse infiltrate consisting of small lymphoid cells without epidermotropism, extending towards the subcutaneous cell tissue, with scarce mitosis. The immunophenotype was CD20(+), CD3(-), and CD5(+).

If a cutaneous B-cell lymphoma is CD5 positive, two possibilities must be considered: a chronic lymphoid leukemia with secondary skin involvement (CD23 positive), or a primary cutaneous mantle B-cell lymphoma (CD23 negative).

In our patient, the immunohistochemistry was negative for CD23 and the diagnosis of leukemia was ruled out through supplementary tests; therefore, we consider that primitive mantle lymphoma should be included in the primary cutaneous B-cell lymphomas.

In contrast with nodal lymphomas, phenotype is not cyclin D1(+), or Bcl-1(+) and there is no chromosomal translocation between sector 13 of the long arm of chromosome 11 and sector 32 of the long arm of chromosome 14 t(11;14)(q13; q32).[77,78]
For all the above, we intend to highlight some facts we deem of interest.
Genetic studies of PCBCL may show monoclonality (Ig light and heavy chain tests) but without evidence of chromosomal translocation t(14,18) characteristic of nodal lymphomas.[79]
Large cell PCBCL of the leg show increased expression of genes associated with cell proliferation cellular: proto-oncogenes *Pim-1*, *Pim-2* and *Myc*; and of genes associated with transcription factors:
Mum1/IRF4 and Oct-2, which demonstrated that they derive from activated B-cells.
In contrast, follicular center lymphomas, when appearing with large cells, have increased SPINK2 expression and a secretion profile similar to germinal center B-cell lymphomas. These investigations suggest different pathogenic mechanisms, and support the WHO-EORTC subdivision.[30,80]
Epstein Barr virus is most frequently found in systemic than in cutaneous immunoblastic lymphomas.[81] Immunosuppresor states (HIV, transplants) modify the course of lymphomas. In case 12 we were able to observe that on the surgical scar of a renal transplant, the patient developed a follicular center PCBC (**Figure 8**). PCR test for Epstein-Barr virus resulted positive in the tumor mass cells. Radiotherapy was indicated, with excellent evolution. The presence of virus did not modify the lymphoma prognosis.[82-84]
Most frequent PCBCL are follicular center lymphomas (56.7 percent), followed by marginal zone (31.4 percent) and leg lymphomas (10.9 percent). These data coincide with international statistics and also with our experience.[9,16,17,38,85] In contrast, marginal zone lymphomas are predominant in Spain.[86]
A study on marginal zone lymphomas communicated four cases localized in head and neck, three with later systemic involvement (one with transformation into large cells, and two cases with Bcl-2 t(14,18) chromosomal translocation.[86] These facts evidence worse prognosis for this location, but do not match our experience (case 16).[87] Another frequently described form is multifocal (72 percent of the cases); in these cases, treatment with chlorambucil is used.[88]
Marking panel makes it possible to establish differences between follicular center and marginal zone lymphomas (**Table 3**).[89]

Figure 8. *Follicular center B-cell lymphoma on renal transplant surgical scar. Positive CRP in tumor tissue for Epstein-Barr virus (Case 12).*

TABLE 3. IMMUNOHISTOCHEMICAL DIFFERENCES BETWEEN PRIMARY CUTANEOUS FOLLICULAR CENTER AND MARGINAL ZONE B-CELL LYMPHOMAS.

	Bcl-6	Bcl-2	CD10	CD21
FC PCBCL *	+	+/-	+	-
MZ PCBCL **	-	+	-	+
*FC: follicular center. **MZ: marginal zone				

Ig heavy chain variable region assessment is useful to evidence monoclonality. In contrast with the light chain immunohistochemical test, this method does not require fresh material.[90,91]
The characteristic chromosomal translocation of nodal lymphomas is established between *Bcl-2* gene in chromosome 18 and the Ig heavy chain-linking region in chromosome 14. Protein Bcl-2 prevents apoptosis. Translocation of its gene leads to overexpression of the protein, thus preventing apoptosis of neoplastic cells and originating greater tumor aggressiveness in nodal lymphomas; this is the main difference with primary cutaneous lymphoma.[92]

Conclusion

The last PCBCL classification took into account clinical, therapeutic, histopatologic, and genetic aspects.[93,94] This global approach optimized dermatologist management of PCBCL and allowed their inclusion in the interdisciplinary group in charge of lymphoma patient follow-up.
Anyhow, the continuous reviewing may produce modifications in a not very distant future.

References

1. Stansfeld AG, Diebold J, Kapanci Y, Kelenyi G, Lennert K, Mioduszewska O, Noel H, Rilke F, Sundstrom C, van Unnik J, Whirght D. Updated Kiel classification for lymphomas. Lancet. 1988; 1:292.
2. The Non-Hodgkin´s Lymphomas Pathologic Classification Project. National Cancer Institute sponsored study of classification of a working formulation for clinical usage. Cancer 982; 49:2112-2135.
3. Harris NL, Jaffe ES, Stein H, Banks PM, et al. A revised European-American classification of lymphoid neoplasms: A proposal from the International Lymphoma Study Group. 1994; 84:1361.
4. Jaffe E, Harris N, Stein H, Vardiman J. WHO classification of tumors: pathology and genetics of tumors of hematopoietic and lymphoid tissues. Lyons, France: IARC Paris; 2001.
5. Sander CA, Flaig MJ, Jaffe ES. Cutaneous manifestations of lymphoma: a clinical guide based on the WHO classification. Clin Lymphoma 2001; 2:86-100.
6. Willemze R, Kerl H, Sterry W, et al. EORTC classification for primary cutaneous lymphomas: a proposal from the Cutaneous Lymphoma Study Group of the European Organization for Research and Treatment of Cancer (EORTC). Blood 1997; 90:354-371.
7. Norton AJ. Classification of cutaneous lymphoma: a critical appraisal of recent proposals. Am J Dermatopathol 1999; 21:279-287.
8. Viglioglia PA. Linfomas cutáneos primitivos (2nd part). Act Terap Dermatol 2002; 74:224-233.
9. Dahbar M, Carbia S, Chain M, Hochman A et al. Linfoma cutáneo de células B centrofolicular: reporte de un caso y actualización. Dermatol Argent 2002; 8:18-21.
10. Russell-Jones R. World Health Organization of hematopoietic and lymphoid tissues: implications for dermatology. J Am Acad Dermatol 2003; 48:93-102.
11. Willemze R, Jaffe ES, Burg G, Cerroni L, Berti E, Swerdlow SH, et al. WHO-EORTC classification for cutaneous lymphomas. Blood 2005; 105:3768-3786.
12. Sander CA, Flaig MJ, Jaffe ES. Cutaneous manifestations of lymphoma: a clinical guide based on the WHO classification. Clin Lymphoma 2001; 2:86-100.
13. Willemze R, Meijer CJLM. EORTC classification for primary cutaneous lymphomas: the best guide to good clinical management. European Organization for Research and Treatment of Cancer.
Am J Dermatopathogy 1999; 21:265-273.
14. Willemze R, Meijer CJLM, Sentis HJ, et al. Primary cutaneous large cell lymphomas of follicular center cell origin. J Am Acad Dermatol 1987; 16:518.
15. Santucci M, Pimpinelli N, Arganini L. Primary cutaneous B-cell lymphoma: a unique type of low-grade lymphoma. Cancer 1991; 67:2311-2326.
16. Berti E, Alessi E, Caputo R, et al. Reticulohistiocytoma of the dorsum. J Am Acad Dermatol 1988; 19:259-272.
17. Crosti A. Micosi fungoide e reticuloistiocitomi cutanei maligni. Minerva Dermat 1951; 26:3-11.
18. Martínez Chabbert P, González Cocorda A, Brusco JE. Linfoma cutáneo de células B centrofolicular (linfoma de Crosti). Dermatol Argent 2005; 11:147-149.
19. Monti J, Bergero A, Lurati C, de la Peña H and cols. Linfoma primario cutáneo fenotipo B de evolution lenta. ¿Por qué no linfoma de Crosti? Estudio de 4 casos. Rev Argent Dermatol 1991; 72:51-56.
20. Cerroni L, Arzberger E, Pütz B, et al. Primary cutaneous follicular center cell lymphoma with follicular growth pattern. Blood 2000; 95:3922-3928.
21. Goodlad JR, Krajewski AS, Batstone PJ, et al. Primary cutaneous follicular lymphoma: a clinicopathologic and molecular study of 16 cases in support of a distinct entity. Am J Surg Pathol 2002;26: 733-741.
22. Grange F, Bekkenk MW, Wechsler J, et al. Prognostic factors in primary cutaneous large B-cell lymphomas: a European multicenter study. J Clin Oncol 2001; 19:3602-3610.
23. Willemze R, Meijer CJLM, Scheffer E, et al. Diffuse large cell lymphomas of follicle center cell origin presenting in the skin: a clinicopathologic and immunologic study of 16 patients. Am J Pathol 1987; 126:325-333.
24. Cerroni L, El-Shabrawi-Caelen L, Pink-Fuches R, et al. Cutaneous spindle-cell lymphoma: a morphologic variant of cutaneous large B-cell lymphoma. Am J Dermatopathol 2000; 22:299-309.
25. De Leval L, Harris NL, Longtine J, Ferry JA, Duncan LM. Cutaneous B-cell lymphomas of follicular and marginal zone types: use of Bcl-6, CD10, Bcl-2, and CD21 in differential diagnosis and classification. Am J Surg Pathol 2001; 25:732-741.
26. Hoefnagel JJ, Vermeer MH, Janssen PM, et al. Bcl-2, Bcl-6 and CD10 expression in cutaneous B-cell lymphoma: further support for a follicle centre cell origin and differential diagnostic significance. Br J Dermatol 2003; 149:1183-1191.
27. Kim BK, Surti U, Pandya AG, Swerdlow SH. Primary and secondary cutaneous diffuse large B-cell lymphomas. Am J Surg Pathol 2003; 27:356-364.
28. Cerroni L, Volkenandt M, Rieger E, Soyer HP, Kerl H. Bcl-2 protein expression and correlation with the interchromosomal (14;18) translocation in cutaneous lymphomas and pseudolymphomas. J Invest Dermatol 1994; 102:231-235.
29. Geelen FAMJ, Vermeer MH, Meijer CJLM, et al. Bcl-2 expression in primary cutaneous large B-cell lymphoma is site-related. J Clin Oncol 1998; 16:2080-2085.
30. Hoefnagel JJ, Dijkman R, Basso K, et al. Distinct types of primary cutaneous large B-cell lymphoma identified by gene expression profiling. Blood 2005;105:3671-3678.
31. Aarts WM, Willemze R, Bende RJ, Meijer CJLM, Pals ST, van Noessel CJ. VH gene analysis of primary cutaneous B-cell lymphomas: evidence for ongoing somatic hypermutation and isotype switching. Blood 1998; 92:3857-3864.
32. Gellrich S, Rutz S, Golembowski S, et al. Primary cutaneous follicle center cell lymphomas and large B-cell lymphomas of the leg descend from germinal center cells: a single cell polymerase chain reaction analysis. J Invest Dermatol 2001; 117:1512-1520.
33. Bekkenk M, Vermeer MH, Geerts ML, et al. Treatment of multifocal primary cutaneous B-cell lymphoma: guidelines of the Dutch Cutaneous Lymphoma Group. J Clin Oncol 1999; 17:2471-2478.
34. Grange F, Petrella T, Beylot-Barry M, et al. Bcl-2 protein expression is the strongest independent prognostic factor of survival in primary cutaneous large B-cell lymphomas. Blood 2004; 103:36623668.
35. Rijlaarsdam JU, Toonstra J, Meijer OW, Noordijk EM, Willemze R. Treatment of primary cutaneous B-cell lymphomas of follicular center cell origin: a clinical follow-up study of 55 patients treated with radiotherapy o polychemotherapy. J Clin Oncol. 1996; 14:549-555.
36. Pimpinelli N, Vallecchi C. Local orthovolt radiotherapy in primary cutaneous B-cell lymphomas: results in a series of 115 patients. Skin Cancer 1999; 14:219-224.

37. Smith BD, Glusac EJ, McNiff JM, et al. Primary cutaneous B-cell lymphoma treated with radiotherapy: a comparison of the European Organization for Research and Treatment of Cancer and the WHO classification systems. J Clin Oncol 2004; 22:634-639.
38. Tous V, Burgos G, Sevinsky L, Sehtman A, et al. Linfoma cutáneo primario de células B centrofolicular. A propósito de un caso con respuesta favorable a la radioterapia. Act Terap Dermatol 2005; 28:94-101.
39. Heinzerling LM, Urbanek M, Funk JO, et al. Reduction of tumor burden and stabilization of disease by systemic therapy with antiCD20 antibody (rituximab) in patients with primary cutaneous B-cell lymphoma. Cancer 2000; 89:1835-1844.
40. Paul T, Radny P, Krober SM, Paul A, Blaheta HJ, Garbe C. Intralesional rituximab for cutaneous B-cell lymphoma. Br J Dermatol 2001; 144:1239-1243.
41. Rijlaarsdam JU, van der Putte SCJ, Berti E, et al. Cutaneous immunocytomas: a clinicopathologic study of 26 cases. Histopathology 1993; 23:119-125.
42. Schmid U, Eckert F, Griesser H, et al. Cutaneous follicular lymphoid hyperplasia with monotypic plasma cells. A clinicopathologic study of 18 patients. Am J Surg Pathol 1995; 19:12-20.
43. Caro WA, Helwing EB. Cutaneous lymphoid Hyperplasia. Cancer 1969; 24:487-502.
44. Evans HL, Winkelmann RK, Banks PM. Differential diagnosis of malignant and benign cutaneous lymphoid infiltrates. A study of 57 cases in which malignant lymphoma had been diagnosed o suspected in the skin. Cancer 1979; 44:699-717.
45. Owen RA, Norton AJ. Cutaneous pseudolymphoma and primary B-cell lymphoma of skin: two disorders o a pathological continuum? J Pathol 1991; 163:169A.
46. Cerroni L, Zöchling N, Pütz B, Kerl H. Infection by Borrelia burgdorferi and cutaneous B-cell lymphoma. J Cutan Pathol 1997; 24:457-461.
47. Wood GS, Kamath NV, Guitart J, et al. Absence of Borrelia burgdorferi DNA in cutaneous B-cell lymphomas from the United States. J Cutan Pathol 2001; 28:502-507.
48. Bailey EM, Ferry JA, Harris NL, Mihm MC, Jacobson JO, Duncan LM. Marginal zone lymphoma (low-grade B-cell lymphoma of mucosa-associated lymphoid tissue type) of skin and subcutaneous tissue: a study of 15 patients. Am J Surg Pathol 1996; 20:1011-1023.
49. De Leval L, Harris NL, Longtine J, Ferry JA, Duncan LM. Cutaneous B-cell lymphomas of follicular and marginal zone types: use of Bcl-6, CD10, Bcl-2, and CD21 in differential diagnosis and classification. Am J Surg Pathol 2001; 25:732-741.
50. Hoefnagel JJ, Vermeer MH, Janssen PM, et al. Bcl-2, Bcl-6 and CD10 expression in cutaneous B-cell lymphoma: further support for a follicle centre cell origin and differential diagnostic ignificance. Br J Dermatol 2003; 149:1183-1191.
51. Gronbaek K, Ralfkiaer E, Kalla J, Skovgaard GL, Guldberg P. Infrequent somatic Fas mutations but no evidence of Bcl-10 mutations or t(11;18) in primary cutaneous MALT-type lymphoma. J Pathol 2003; 201:134-140.
52. Hallermann C, Kaune KM, Gesk S, et al. Molecular cytogenetic analysis of chromosomal breakpoints in the IGH, MYC, BCL6 and MALT1 gene loci in primary cutaneous B-cell lymphomas. J Invest Dermatol. 2004; 123:213-219.
53. Li C, Inagaki H, Kuo TT, Huo S, Okabe M, Eimoto T. Primary cutaneous marginal zone B-cell lymphoma: a molecular and clinicopathologic study of 24 Asian cases. Am J Surg Pathol 2003; 27: 1061-1069.
54. Zenahlik P, Pink-Fuches R, Kapp KS, Kerl H, Cerroni L. Therapy of primary cutaneous B-cell lymphomas. Hautarzt 2000; 51:19-24.
55. Soda R, Constanzo A, Cantonetti M, Orlandi A, Bianchi L, Chimenti S. Systemic therapy of primary cutaneous B-cell lymphoma, marginal zone type, with rituximab, a chimeric anti-CD20 monoclonal antibody. Acta Derm Venereol 2001; 81:207-208.
56. Wiltshaw E. The natural history of extramedullary plasmacytoma and its relation to solitary myeloma of bone and myelomatosis. Medicine 1976; 553:217.
57. Torne R, Su WPD, Winkelmann RK, Smolle J, Kerl H. Clinicopathologic study of cutaneous plasmacytoma. Int J Dermatol 1990; 29:562-566.
58. Chang YT, Wong CK. Primary cutaneous plasmacytomas. Clin Exp Dermatol 1994; 19:177.
59. Vermeer MH, Geelen FAMJ, van Haselen CW, et al. Primary cutaneous large B-cell lymphomas of the legs: a distinct type of cutaneous B-cell lymphoma with an intermediate prognosis. Arch Dermatol 1996; 132:1304-1308.
60. Goodlad JR, Krajewski AS, Batstone PJ, et al. Primary cutaneous diffuse large B-cell lymphoma: prognostic significance and clinicopathologic subtypes. Am J Surg Pathol 2003; 27:1538-1545.
61. Grange F, Petrella T, Beylot-Barry M, et al. Bcl-2 protein expression is the strongest independent prognostic factor of survival in primary cutaneous large B-cell lymphomas. Blood 2004; 103:36623668.
62. Paulli M, Viglio A, Vivenza D, et al. Primary cutaneous large B-cell lymphoma of the leg: histogenetic analysis of a controversial clinicopathologic entity. Hum Pathol 2002; 33:937-943.
63. Mao X, Lillington D, Child FJ, Russell-Jones R, Young B, Whittaker S. Comparative genomic hybridization analysis of primary cutaneous B-cell lymphomas: identification of common genomic alterations in disease pathogenesis. Genes Chromosomes Cancer 2002; 35:144-155.
64. Child FJ, Scarisbrick JJ, Calonje E, Orchard G, Russell-Jones R, Whittaker SJ. Inactivation of tumor suppressor genes p15(INK4b) and p16(INK4a) in primary cutaneous B cell lymphoma. J Invest Dermatol 2002; 118:941-948.
65. Hallermann C, Kaune K, Siebert R, et al. Cytogenetic aberration patterns differ in subtypes of primary cutaneous B-cell lymphomas. J Invest Dermatol 2004; 122:1495-1502.
66. Hallermann C, Kaune KM, Gesk S, et al. Molecular cytogenetic analysis of chromosomal breakpoints in the IGH, MYC, BCL6 and MALT1 gene loci in primary cutaneous B-cell lymphomas. J Invest Dermatol 2004; 123:213-219.
67. Brogan BL, Zic JA, Kinney MC, Hu JY, Hamilton KS, Greer JP. Large B-cell lymphoma of the leg: clinical and pathologic characteristics in a North American series. J Am Acad Dermatol 2003; 49:223-228.
68. Colomo L, Loong F, Rives S, et al. Diffuse large B-cell lymphomas with plasmablastic differentiation represent a heterogeneous group of disease entities. Am J Surg Pathol 2004; 28:736-747.
69. Sander CA, Kaudewitz P, Kutzner H, et al. T-cell rich B-cell lymphoma presenting in the skin. A clinicopathologic analysis of six cases. J Cutan Pathol 1996; 23:101-108.
70. Li S, Griffin CA, Mann RB, Borowitz MJ. Primary cutaneous T-cell rich B-cell lymphoma: clinically distinct from its nodal counterpart? Mod Pathol 2001; 14:10-13.
71. Perniciaro C, Winkelmann RK, Daoud MS, Su WPD. Malignant angioendotheliomatosis is an angiotropic intravascular lymphoma: immunohistochemical, ultrastructural and molecular genetic studies. Am J Dermatopathol 1995; 17:242-248.
72. Ferreri AJM, Campo E, Seymour JF, et al. Intravascular lymphoma: clinical presentation, natural history, management and prognostic factors in a series of 38 cases with special emphasis on the "cutaneous variant." Br J Haematol 2004; 127:173-183.
73. Rubin MA, Cossman J, Freter CE, Azumi N. Intravascular large cell lymphoma coexisting within hemangiomas of the skin. Am J Surg Pathol 1997; 21:860-864.
74. Kobayashi T, Munakata S, Sugiura H, et al. Angiotropic lymphoma: proliferation of B-cells in the capillaries of cutaneous angiomas. Br J Dermatol 2000; 143:162-164.

75. Bertero M, Novelli M, Fierro M T, Bernengo M G. Mantle zone lymphoma: An imnunohistochemicalstudy of skin lesions. J Am Acad Dermatol 1994; 30:23.
76. Greets M L, Busschots A. Mantle-cell lymphomas of the skin. Dermatol Clin 1994;12;409.
77. Kurtin P J. Mantle cell lymphoma. Adv Anat Pathol 1998;5:376-398.
78. Campo E, Raffeld M, Jaffe ES. Mantle-cell lymphoma. Semin Hematol 1999; 36:115-117.
79. Grange F, Hedelin G, Joly P, et al. Prognostic factors in primary cutaneous lymphomas other than mycosis fungoides and the Sezary syndrome: the French study group on cutaneous lymphomas. Blood 1999; 93:3637-3642.
80. Dijkman R, Tensen DP, Jordanova ES, Knijnenburg J, et al. Arraybased comparative genomic hybridization analisys reveals recurrent chromosomal alterations and prognostic parameters in primary cutaneous large B-cell lymphoma. J Clin Oncol 2006; 24:296-305.
81. Gaidano G, Carbone A, Dalla Favera R. Patogenesis of AIDS-related lymphomas. Am J Pathol 1998; 152:623-630.
82. Copur MS, Deshpande A, Mleczko K, Norvell M, Hrnicek GJ, Woodward S, Frankforter S, Mandolfo N, Fu K, Chan WC. Full clinical recovery after topical acyclovir treatment of Epstein Barr virus associated cutaneous B-cell lymphoma in patient with mycosis fungoides. Croat Med J 2005; 46:458-462.
83. Verma S, Frambach GE, Seilstad KH, Nuovo G, Porcu P, Magro CM. Epstein- Barr virus associated B-cell lymphoma in the setting of iatrogenic immune dysregulation presenting initially in the skin. J Cutan Pathol 2005; 32:474-483.
84. Villoldo V, Forero O, Dionisio M, Anaya J, Magariños G, Pizzariello G. Linfoma B en un paciente HIV positivo. Dermatol Argent 2004; 4:284-290.
85. Zinzani PL, Quaglio P, Pimpinelli N, Berti E, et al. Prognostic factors in primary cutaneous B-cell lymphoma: the Italian Study Group for Cutaneous Lymphomas. J Clin Oncol 2006; 24:1376-1382.
86. Servitje Bedate O. Linfomas cutáneos primarios de células B. Concepto, características clínico-patológicas y clasification. Dermatología 2001; 14:163-168.
87. Servitje O, Gallardo F, Estrach T, Pujol RM, et al. Primary cutaneous marginal zone B-cell lymphoma: a clinical, histopathological, inmunophenotypic and molecular genetic study of 22 cases. Br J Dermatol 2002; 147:1147-1158.
88. Hoefnagel JJ, Vermeer MH, Jansen PM, van Voorst Vader PC, et al. Primary cutaneous marginal zone B-cell lymphoma: clinical and therapeutic features in 50 cases. Arch Dermatol 2005; 141:1139-1145.
89. De Leval L, Harris NL, Longtine J, Ferri JA, Duncan LM. Cutaneous B-cell lymphomas of follicular and marginal zone types: use of Bcl-6, CD10, bcl-2, and CD21 in differential diagnosis and classification. Am J Surg Pathol 2001; 25:732-741.
90. Child FJ, Woolford AJ, Calonje E, Russell-Jones R, Whittaker SJ. Molecular analysis of the immunoglobulin heavy chain gene in the diagnosis of primary cutaneous B cell lymphoma. J invest Dermatol 2001; 117:984-989.
91. Alaibac M, Belloni-Fortina A, Mori M, Pigozzi B, Peserico A, Pimpinelli N. Inmunoglobulin heavy chain variable region family expression in primary cutaneous follicle centre cell lyphomas. Br J Dermatol 2001; 144:862-865.
92. Kim BK, Surti U, Pandya, AG, Swerdlow SH. Primary and secondary cutaneous diffuse large B-cell lymphomas: a multiparameter analysis of 25 cases including fluorescence in situ hybridization for t(14;18) translocation. Am J Pathol 2003; 27:356-364.
93. Slater DN. The new World Health Organization – European Organization for Research and Treatment of Cancer classification for cutaneous lymphomas: a practical marriage of two giants. Br J Dermatol 2005; 153:874-880.
94. Burg G, Kempf W, Cozzio A, Feit J, et al. WHO/EORTC classification of cutaneous lymphomas 2005: histological an molecular aspects. J Cutan Pathol 2005; 32:647-674.